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# USF1 gene polymorphisms may associate with the efficacy and safety of chemotherapy based on paclitaxel and prognosis in the treatment of ovarian cancer

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This study was supposed to investigate the correlation between the functional single nucleotide polymorphisms (SNPs) (rs2516839 and rs3737787) in USF1 gene and the efficacy and safety of paclitaxel-based chemotherapy and prognosis in the treatment of ovarian cancer (OC). In total 100 OC patients were selected and divided into the sensitive group and the resistant group according to the tumor response to paclitaxel-based chemotherapy after surgery, and the incidence of observed and recorded toxic reaction. Polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method was applied to test the polymorphisms of rs2516839 and rs3737787 in USF1 gene after extraction of DNA. The correlation between USF1 gene polymorphisms and paclitaxel-based chemotherapy resistance was analyzed using Logistic regression analysis. Stratified analysis was used to test the incidence of toxic reaction in OC patients. Cox proportional hazard model was adapted to make a multiple-factor survival analysis. Significant differences exhibited in the genotype and the allele frequencies of rs2516839 between the sensitive and resistant groups, which showed no obvious difference in the genotype and allele frequencies of rs3737787. OC patients carrying the GA+AA genotype had higher incidence of serious toxic reaction than those carrying the GG genotype. Physical status score, tumor type, maximum tumor diameter and rs2516839 polymorphism in USF1 gene may associate with the efficacy and safety of paclitaxel-based chemotherapy and prognosis in the treatment of OC.

Key words: drug resistance, gene polymorphisms, Paclitaxel-based chemotherapy regimen, prognosis, ovarian cancer, rs2516839 (G>A), rs3737787(C>T), USF1 gene

Ovarian cancer (OC), the fourth commonest gynecologic cancer, has been the third leading reason of death caused by gynecologic malignancy among women [1]. OC patients are typically diagnosed at a late stage, with a 5-year survival rate of less than 30% [2]. Symptoms of OC are indeterminate, including abdominal discomfort, cacochylia, and abdominal distension [3]. The diffuse spread of OC is considered to arise through the peritoneal circulation, related to ascites formation [2].

Since the mid-1990s, maximal surgical debulking combined with platinum-based chemotherapy has been the standard approach in treating advanced OC, which lead to a higher response rate of approximately 75% in OC patients [4]. However, the rates of palindromia and mortality are still high (5-year survival rate: 20–40%) and most patients eventually recrudesce because of drug resistance [5]. Identifying new powerful medication and establishing better chemotherapy regime are extremely urgent. Recently, the chemotherapy based on paclitaxel-platinum, which includes surgical staging and resection, has been identified as one of the universal therapeutic methods in treating advanced epithelial OC [6]. However, 75% patients eventually relapse with drug resistant disease within 18 months [7].

As one of the most important factors for the treatment of failure or death in patients with OC, the mechanism of paclitaxel resistance is complicated and not yet completely expounded [8]. A study has demonstrated that changes involved in glycolysis (HK1, ADH1A, and ENO3), oxidative stress (MAOA, UGT1A6, and CYBA), mRNA and protein (translation factors and multiple ribosomal genes) synthesis, leukocyte transendothelial migration pathways and glutathione metabolism are the major causes of paclitaxel resistance [9]. Another study reported that paclitaxel resistance identified changes in pathways associated with mRNA and protein synthesis, and multiple ribosomal genes (RPS20, RPL26, RPL10A, RPL39, RPL7, and RPL34) and translation factors [7]. Recently, the correlation between the mutation of TEKT4 gene and paclitaxel resistance has been demonstrated [10]. Preventing or delaying the development of paclitaxel resistance may be greatly conductive to improve the clinical outcome of cancer patients.

The Upstream stimulatory factor 1 (*USF1*) is considered to be one of the ubiquitous transcription factors of the bzHLH (leucine zipper-basic-helix-loop-helix) family [11]. As an important regulator of enormous genes, *USF1* is involved in thrombotic complications of the coronary plaque, stress and immune responses, cell cycle and differentiation, and lipid and glucose metabolism [12]. Familial combined hyperlipidemia and one of its component traits had been reported to correlate with *USF1* gene polymorphisms in many studies and were also analyzed in the context of diabetes mellitus type II, metabolic syndrome and CAD [13].

Evidence has demonstrated that protein expressions of *JAK2*, *HSP*, *MSH2*, *WNT-1* and *FZD-1* gene are associated with paclitaxel-based chemotherapy resistance of OC patients [14]. However, little attention has been paid to the relationship between *USF1* gene polymorphisms and paclitaxel-based chemotherapy resistance of OC patients. Thus, in this study, we aimed to explore the correlations of the functional SNPs (rs2516839 and rs3737787) in *USF1* gene with the efficacy and safety of paclitaxel-based chemotherapy and prognosis in treating patients with OC.

## Materials and methods

Study subjects. Between February 2009 and March 2011, a total of 122 patients (Han nationality, age: 27-78 years, median age: 51 years) diagnosed with OC and treated in The First Clinical Medical College of China Three Gorges University, were divided into stage II (n=35), stage III (n=64) and stage IV (n=23) (2010) [15]. Pathologic types were categorized into serous cystadenocarcinoma based on the International Federation of Gynecology and Obstetrics (FIGO) staging (n=68), mucinous cystadenocarcinoma (n=16), endometrioid carcinoma (n=19) and other epithelial cancers (n=19). According to the diagnostic criteria recommended by the World Health Organization (WHO) [16], the tumors were classified into well differentiated (n=63), moderately differentiated (n=38), and poorly differentiated (n=21). The inclusion criteria were as follows: (1) Patients pathologically diagnosed as primary OC; (2) OC patients who never received chemotherapy and hormone therapy; (3) OC patients who were continuously treated with paclitaxel-based chemotherapy for at least 6 cycles; (4) OC patients with no history of other cancers and concomitant malignant tumor. Importantly, this study was performed with the approval of the Ethics committee of The First Clinical Medical College of China Three Gorges University, which also strictly followed the ethical principles involving human subjects as formulated

in the Helsinki Declaration. Informed consent was obtained from every participant or their legal guardians.

Chemotherapy regimens and efficacy evaluation. The chemotherapy regimen was illustrated in a combination of paclitaxel (175 mg/m2, 3-hour intravenous infusion) and carboplatin (AUC=5-7.5) once every 3 weeks for a total of 6-8 cycles. Tumor responses were categorized as complete response (CR) (n=28), partial response (PR) (n=61), stable disease (SD) (n=25), or progressive disease (PD) (n=8) according to the Clinical Curative Effect Criteria for Solid Tumors stipulated by WHO [16], the total response rate (RR)=CR+PR. Patients were divided into the sensitive group (CR+PR, n=89) and the resistant group (SD+PD, n=33) according to the curative effect of paclitaxel-based chemotherapy. According to the toxic reaction, all patients with OC were graded (grade 0-4) in strict accordance with the Response Evaluation Criteria in Solid Tumors (RECIST) guidelines of World Health Organization (WHO) [16], including anaphylaxis, peripheral nerve toxicity, vomiting, diarrhea, and aleucocytosis etc.

**DNA extraction**. After obtaining informed consent, 5 ml of venous blood were taken from patients and put into the EP tube for 15–25 minutes at 37 °C, oscillated and mixed by adding 1 ml lysate, and centrifuged by 12000 r/min for 5 minutes after putting in room-temperature for 15 minutes. DNA extraction was carried out with a genomic DNA extraction kit (Promega, US). The concentration (100±20 mg/L) of the DNA was assessed using NANODROP2000 (Thermo, Massachusetts, US), and the purity was assessed by the optical density (OD) measurement at 260 nm. The average purity (OD260/OD280) of the samples was required to be between 1.6 and 1.9, and DNA samples were kept frozen at -20 °C.

Genotyping. Primers were synthesized by Le Jin Biotech (Nanjing) Co., Ltd. For rs2516839, the forward primer was 5'-ACGTTGGATGCTGGTCCTTTTTTGGAGGTC-3' and reverse primer was 5'-ACGTTGGATGTCTACCAGGACT-TAGCACTC-3' [13]. For rs3737787, the forward primer was 5'-ACGTTGGATGAGAGGAGCACAAGGGCCCA-3' and reverse primer was 5'-ACGTTGGATGCAGTGGTGT-GAAACACACAA-3' [17]. The experiment procedures were performed using polymerase chain reaction (PCR) kits (Univ Biological Technology Co., Ltd, Shanghai). The PCR reaction system was set up in a total volume of 20 µL with a mixture of 1 µL DNA samples, 1 µL forward primer (10 pmol/ml), 1 µL reverse primer (10 pmol/ml), 7 µL deionized water and 10 µL PCR. The PCR reaction conditions were arranged as follows: pre-denaturalization at 95 °C for 5 minutes; then 35 cycles of denaturalization for 1 minute at 95 °C, anneal at 65 °C for 30 seconds, extension at 72 °C for 1 minute; and the final step was elongation for 10 minutes at 72 °C. In the reaction mixtures 4µL of amplification products and 1µL of bromophenol blue solution were added, which then were subjected to electrophoresis in 3% agarose gel. Running time in  $0.5 \times \text{TBE}$  buffer was 15 minutes (75 V) and amplification band was visualized under UV light. PCR-amplified products of rs2516839 and rs3737787 were digested with restriction enzymes NCOI at 50 °C and NlaIII at 37 °C (NEB Company, US, Beijing branch). Digestion products were resolved by electrophoresis on a 3% agarose gels (110 V, 40min) and visualized under UV light. Ten percent of PCR products were randomly selected and sent to Shanghai Sangon Biological Engineering Technology Co., Ltd and the results of polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) were verified by completely sequencing the PCR products from both directions.

**Follow-up.** A regular follow-up of 1~5 years for 100 OC patients (follow-up rate 88.52%) was performed by outpatient service, return visit, follow-up letters and telephone interview, including the results of last examination, gynecological examination, imaging examination, tumor markers examination and physical status score etc. The overall 1-year, 3-year and 5-year survival rate was 88.89%, 69.44% and 37.96%. Follow-up start time was defined as the first month after treatment. Follow-up appointments were scheduled once in 3 months within the first year and 1–2 times per year later until the occurrence of death.

Statistical analysis. SPSS 20.0 integrated software (SPSS Inc. IBM, Chicago, IL, USA) was applied for the statistical analysis. Continuous data was shown as mean ± standard deviation  $(\pm s)$ . The t-test was used for the comparison of two groups, and one-way analysis of variance (ANOVA) for comparison among multi-group. Welch's t-test was explored for the data that did not meet the assumption of equal variances. Pairwise comparisons were performed using the Least Significant Difference (LSD) test (for homogeneity of variance) or Tamhane's test (for heterogeneity of variance). The differences in genotype and allele frequencies among groups were identified using Pearson's Chi square test. Logistic regression analysis was conducted for the relationship between USF1 gene polymorphisms and paclitaxelbased chemotherapy resistance. Hardy-Weinberg equilibrium was measured by the chi-square test. Survival was calculated by the Kaplan-Meier (KM) method and the Cox proportional hazard model was adapted to make a multiplefactor analysis. All statistical tests were two-sided, with p<0.05 being considered statistically significant.

## Results

Baseline characteristics of OC patients between the sensitive and resistant groups. A total of 122 OC patients were categorized into a sensitive group (n=89; mean age:  $51.4\pm12.9$  years; mean weight:  $54.1\pm8.9$  kg) and a resistant group (n=33; mean age:  $53.6\pm12.2$  years; mean weight:  $53.0\pm7.7$  kg). As shown in Table 1, the baseline characteristics of the 122 patients revealed that no significant difference existed in the age and weight between the sensitive and resistant groups (all p>0.05). Physical status score, FIGO stage, pathologic types and maximum tumor diameter also exhibited no notable difference between these two groups (all p>0.05).

The incidences of toxic reaction in OC patients between the sensitive and resistant groups. After the treatment of paclitaxel-based chemotherapy, no toxicity-related death was found in the sensitive and resistant groups, while the severe toxic reactions with the highest incidence (grade 3–4) were observed. There were 32 cases of anaphylaxis, 35 cases of peripheral nerve toxicity, 33 cases of vomiting, 38 cases of diarrhea and 37 cases of aleucocytosis in the two groups, while no significant difference in the incidence of the toxic reactions was identified between the sensitive and the resistant group (p>0.05) (Table 2).

Table 1. Baseline characteristics of OC patients between the sensitive and resistant groups.

Baseline characteristics	Resistant group	Sensitive group	p-value
Dasenne characteristics	(n=33)	(n=89)	p-value
Age (years)	()	()	
<45	6	19	0.890
45-60	13	36	
>60	14	34	
Mean weight(kg)	53.0±7.7	54.1±8.9	0.531
Physical status			
0–1 point	24	74	0.156
2 points	9	15	
Pathologic types			
Serous adenocarcinoma	13	55	0.133
Endometrioid carcinoma	9	10	
Clear-cell carcinoma	3	9	
Mucinous adenocarcinoma	6	10	
Undifferentiated carcinoma	2	5	
Maximum tumor diameter (cm)			
<1	12	42	0.520
1–2	14	29	
>2	7	18	
Tumor stage			
II	7	28	0.274
III	17	47	
IV	9	14	

Table 2. The incidences of toxic reaction in OC patients between the sensitive and resistant groups.

Toxic reaction	Resistant group (n=33)	Sensitive group (n=89)	p-value
Anaphylaxis	3/12	10/20	0.492
(0-2/3-4)	5/12	10/20	0.472
Peripheral nerve toxicity	2/16	9/18	0.656
(0-2/3-4)	2/10	5/18	0.050
Vomiting	1/4	9/19	0.586
(0-2/3-4)	1/4	9/19	0.580
Diarrhea	3/8	9/18	0.715
(0-2/3-4)	5/8	5/18	0.715
Aleucocytosis	6/7	5/19	0.108
(0-2/3-4)	0/7	5/19	0.108

The distribution of the allele and genotype frequencies of USF1 gene polymorphisms. The allele frequencies of the rs2516839 and rs3737787 in USF1 gene were in a state of equilibrium according to the Hardy-Weinberg equilibrium and the population was well represented by samples. As shown in Table 3, the distribution of allele and genotype frequencies of rs2516839 were significantly different between the sensitive and resistant groups (both p<0.05), thus the rs2516839 polymorphisms in USF1 gene might be associated with increase of paclitaxel resistance of OC (GA vs. GG, OR=2.246, 95%CI=1.252-4.028, p<0.05; AA vs. GG, OR=2.246, 95%CI=1.252-4.028, p<0.05; GA+AA vs. GG, OR=2.246, 95%CI=1.252-4.028, p<0.05; A vs. G, OR=7.768, 95%CI=1.147-2.725, p<0.05). However, there was no evident difference in terms of the genotype and allele frequencies of the rs3737787 between the sensitive and resistant groups (all p>0.05).

Correlations between the USF1 gene polymorphisms and the clinicopathological features of OC patients. As shown in Table 4, stratified analysis of clinical data (age, mean age and mean weight) was performed, indicating that there was no statistical significance in the difference between frequencies of GG genotype and GA+AA genotype of rs2516839 in USF1 gene (p>0.05). Based on the stratified analysis of physical status score, tumor pathology and maximum tumor diameter, the frequencies of the GG genotype of rs2516839 was notably different with the GA+AA genotype (p<0.05). Compared with the patients with a physical status score of 2 points and maximum tumor diameter less than or equal to 2 cm, the frequencies of GA+AA genotype among OC patients with a physical status score of 0-1 point and maximum tumor diameter larger than 2 cm was significantly higher, the proportion of patients with ovarian serous adenocarcinomas in OC patients carrying the GG genotype was notably increased (p<0.05). However, there was no significant difference in the frequencies between the CC genotype and CT+TT genotype of rs3737787 in *USF1* gene according to the stratified analysis of age, mean age, mean weight, physical status score, pathological type of tumors and maximum tumor diameter (p>0.05).

Associations of the USF1 gene polymorphisms with the incidence of toxic reactions among OC patients. As can be seen in Table 5, a total of 122 OC patients were stratified based on the toxic reactions to paclitaxel-based chemotherapy. Grade 3–4 was regarded as severe toxic reaction. The incidences of the serious toxic reactions in patients carrying GA+AA genotype of rs2516839 in USF1 gene were obviously higher than in the patients carrying GG genotype (p<0.05). There was no significant difference between the frequencies of the distribution of CC genotype and CT +TT genotype of rs3737787 in USF1 gene based on the stratified analysis of anaphylaxis, peripheral nerve toxicity, vomiting, diarrhea and aleucocytosis (p>0.05).

Relationship between the USF1 gene polymorphism and prognosis of OC patients. The results based on Kaplan-Meier curves (Figures 1 and 2) indicated that the 5-year cumulative survival rates of the OC patients were 37.96%, while the cumulative survival rates of OC patients carrying GG and GA+AA genotype were 55.32% and 20.00% respectively. The difference in the cumulative survival rates of the OC patients carrying GG genotype and OC patients carrying GA+AA genotype was statistically significant (p<0.05). The results of the Cox multiple regression analysis revealed that the physical status score (OR=1.909), tumor pathology (OR=2.650), maximum tumor diameter (OR=1.729) and rs2516839 (OR=1.729) were independent risk factors for the prognosis of OC patients. Physical status score (2 points), maximum tumor diameter (larger than 2 cm), serous adenocarcinoma and GA+AA genotype of rs2516839 were the risk factors for prognosis of OC patients (all p<0.05) (Table 6).

SNPs	Resistant group	Sensitive group		OD	050/01
	(n=33)	(n=89)	p-value	OR	95%CI
rs2516839 (G>A)					
GG	7 (21.2%)	40 (44.9%)			Reference
GA	8 (24.2%)	13 (14.6%)	0.033	3.516	1.068-11.580
AA	18 (54.5%)	36 (40.4%)	0.015	3.420	1.220-9.540
GA+AA	26 (78.8%)	49 (55.1%)	0.005	3.840	1.450-10.200
G allele	22 (33.3%)	93 (52.2%)			Reference
A allele	44 (66.7%)	85 (47.8%)	0.007	2.240	1.240-4.050
s3737787 (C>T)					
CC	17 (51.5%)	43 (48.3%)			Reference
CT	10 (30.3%)	28 (31.5%)	0.828	1.107	0.443-2.764
ГТ	6 (18.2%)	18 (20.2%)	0.757	1.186	0.402-3.498
CT+TT	16 (48.5%)	46 (51.7%)	0.753 1.137		0.511-2.528
C allele	44 (66.7%)	114 (64.0%)			Reference
Γ allele	22 (33.3%)	64 (36.0%)	0.703	1.123	0.618-2.039

Note: SNPs, functional single nucleotide polymorphisms;

Clinical features	rs251683	rs2516839 (G>A)	- p-value OR	OD	R 95%CI -	rs373778	87 (C>T)	– p-value	OR	95%CI
Chinical leatures	GG	GA+AA		OK	95%CI	CT+TT	CC			
Age (years)										
<45	9 (19.1%)	16 (21.3%)	0.846	0.873	0.351-2.176	12 (19.4%)	13 (21.7%)	0.741	0.867	0.360-2.092
45-60	18 (38.3%)	31 (41.3%)				27 (43.5%)	22 (36.7%)			
>60	20 (42.6%)	28 (37.3%)				23 (37.1%)	25 (41.7%)			
Mean weight (kg)	54.7±8.8	53.2±8.3	0.344	1.010	1.020-1.330	53.0±8.7	54.6±8.3	0.301	1.21	0.75 - 1.50
Physical										
Status score										
0-1	31 (66.0%)	67 (89.3%)	0.002	4.323	1.672-11.170	44 (71.0%)	54 (90.0%)	0.083	0.442	0.173-1.129
2	16 (34.0%)	8 (10.7%)				18 (29.0%)	6 (10.0%)			
Pathology										
Serous adeno carcinoma	30 (63.8%)	38 (50.7%)	0.030	1.718	0.813-3.630	34 (54.8%)	34 (56.7%)	0.429	0.929	0.454-1.898
Endometrioid carcinoma	11(25.5%)	8 (10.7%)				8 (12.9%)	11(18.3%)			
Clear-cell carcinoma	2 (4.3%)	10 (13.3%)				9 (14.5%)	3 (5.0%)			
Mucinous adeno carcinoma	3 (6.4%)	13 (17.3%)				7 (11.3%)	9 (15.0%)			
Undifferentiated carcinoma	1 (2.1%)	6 (8.0%)				4 (6.5%)	3 (5.0%)			
Maximum tumor diameter (cm)										
<1	28 (59.6%)	26 (34.7%)	0.003	2.777	1.309-5.893	26 (41.9%)	28 (46.7%)	0.718	0.825	0.404-1.688
1–2	8 (17.0%)	35 (46.7%)				24 (38.7%)	19 (31.7%)			
>2	11 (23.4%)	14 (18.7%)				12 (19.4%)	13 (21.7%)			
Tumor stage										
II	20 (42.6%)	15 (20.0%)	0.016	2.960	1.320-6.660	16 (25.8%)	19 (31.7%)	0.772	0.7506	0.342-1.650
III	22 (46.8%)	42 (56.0%)				34 (54.8%)	30 (50.0%)			
IV	5 (10.6%)	18 (24.0%)				12 (19.4%)	11 (18.3%)			

Table 4. Correlation between the USF1	gene polymorphisms and the clinic	opathological features of OC patients.

Table 5. Association of the USF1	gene	polymor	phisms with the incie	dence of toxic reactions	among OC patients.

rs25168	39 (G>A)		OD	05% CI	rs37377	87 (C>T)		OD	050/ 01
oxic reactions GG GA	GA+AA	- p-value	OK	95%CI	CT+TT	CC	- p-value	OK	95%CI
8 (47.1%)	5 (17.9%)	0.036	4.089	1.052-15.890	6 (30.0%)	7 (28.0%)	0.883	1.102	0.3018-4.025
9 (52.9%)	23 (82.1%)				14 (70.0%)	18 (72.0%)			
6 (60.0%)	5 (20.0%)	0.021	6.000	1.211-29.740	8 (47.1%)	3 (16.7%)	0.053	4.440	0.931-21.230
4 (40.0%)	20 (80.0%)				9 (52.9%)	15 (83.3%)			
7 (58.3%)	3 (14.3%)	0.008	8.400	1.570-44.900	4 (21.1%)	6 (42.9%)	0.180	0.356	0.077-1.640
5 (41.7%)	18 (85.7%)				15 (78.9%)	8 (57.1%)			
8 (57.1%)	4 (16.7%)	0.010	6.667	1.476-30.120	7 (38.9%)	5 (25.0%)	0.358	1.909	0.477-7.641
6 (42.9%)	20 (83.3%)				11 (61.1%)	15 (75.0%)			
6 (60.0%)	5 (18.5%)	0.014	6.600	1.339-32.530	4 (19.0%)	7 (43.8%)	0.103	0.302	0.070-1.317
4 (40.0%)	22 (81.5%)				17 (81.0%)	9 (56.3%)			
	GG 8 (47.1%) 9 (52.9%) 6 (60.0%) 4 (40.0%) 7 (58.3%) 5 (41.7%) 8 (57.1%) 6 (42.9%) 6 (60.0%)	8 (47.1%)       5 (17.9%)         9 (52.9%)       23 (82.1%)         6 (60.0%)       5 (20.0%)         4 (40.0%)       20 (80.0%)         7 (58.3%)       3 (14.3%)         5 (41.7%)       18 (85.7%)         8 (57.1%)       4 (16.7%)         6 (42.9%)       20 (83.3%)         6 (60.0%)       5 (18.5%)	GG         GA+AA         p-value $8 (47.1\%)$ $5 (17.9\%)$ $0.036$ $9 (52.9\%)$ $23 (82.1\%)$ $6 (60.0\%)$ $5 (20.0\%)$ $0.021$ $4 (40.0\%)$ $20 (80.0\%)$ $0.021$ $4 (40.0\%)$ $20 (80.0\%)$ $0.008$ $7 (58.3\%)$ $3 (14.3\%)$ $0.008$ $5 (41.7\%)$ $18 (85.7\%)$ $0.010$ $6 (42.9\%)$ $20 (83.3\%)$ $0.014$	GG         GA+AA         p-value         OR $8 (47.1\%)$ $5 (17.9\%)$ $0.036$ $4.089$ $9 (52.9\%)$ $23 (82.1\%)$ $0.021$ $6.000$ $6 (60.0\%)$ $5 (20.0\%)$ $0.021$ $6.000$ $4 (40.0\%)$ $20 (80.0\%)$ $0.008$ $8.400$ $7 (58.3\%)$ $3 (14.3\%)$ $0.008$ $8.400$ $5 (41.7\%)$ $18 (85.7\%)$ $0.010$ $6.667$ $6 (42.9\%)$ $20 (83.3\%)$ $0.014$ $6.600$	GG         GA+AA         p-value         OR         95%CI           8 (47.1%)         5 (17.9%)         0.036         4.089         1.052–15.890           9 (52.9%)         23 (82.1%)         0.021         6.000         1.211–29.740           4 (40.0%)         20 (80.0%)         0.008         8.400         1.570–44.900           5 (41.7%)         18 (85.7%)         0.010         6.667         1.476–30.120           6 (60.0%)         5 (18.5%)         0.014         6.600         1.339–32.530	GG         GA+AA         p-value         OR         95%CI         CT+TT $8 (47.1\%)$ $5 (17.9\%)$ $0.036$ $4.089$ $1.052-15.890$ $6 (30.0\%)$ $9 (52.9\%)$ $23 (82.1\%)$ $14 (70.0\%)$ $14 (70.0\%)$ $6 (60.0\%)$ $5 (20.0\%)$ $0.021$ $6.000$ $1.211-29.740$ $8 (47.1\%)$ $4 (40.0\%)$ $20 (80.0\%)$ $0.021$ $6.000$ $1.211-29.740$ $8 (47.1\%)$ $7 (58.3\%)$ $3 (14.3\%)$ $0.008$ $8.400$ $1.570-44.900$ $4 (21.1\%)$ $5 (41.7\%)$ $18 (85.7\%)$ $15 (78.9\%)$ $15 (78.9\%)$ $8 (57.1\%)$ $4 (16.7\%)$ $0.010$ $6.667$ $1.476-30.120$ $7 (38.9\%)$ $6 (42.9\%)$ $20 (83.3\%)$ $11 (61.1\%)$ $6.600$ $1.339-32.530$ $4 (19.0\%)$	GG         GA+AA         p-value         OR         95%CI $CT+TT$ CC           8 (47.1%)         5 (17.9%)         0.036         4.089         1.052–15.890         6 (30.0%)         7 (28.0%)           9 (52.9%)         23 (82.1%)         14 (70.0%)         18 (72.0%)           6 (60.0%)         5 (20.0%)         0.021         6.000         1.211–29.740         8 (47.1%)         3 (16.7%)           4 (40.0%)         20 (80.0%)         9 (52.9%)         15 (83.3%)         9 (52.9%)         15 (83.3%)           7 (58.3%)         3 (14.3%)         0.008         8.400         1.570–44.900         4 (21.1%)         6 (42.9%)           5 (41.7%)         18 (85.7%)         0.010         6.667         1.476–30.120         7 (38.9%)         5 (25.0%)           6 (42.9%)         20 (83.3%)         0.014         6.600         1.339–32.530         4 (19.0%)         7 (43.8%)	GGGA+AAp-valueOR95%Cl $CT+TT$ CCp-value8 (47.1%)5 (17.9%)0.0364.0891.052–15.8906 (30.0%)7 (28.0%)0.8839 (52.9%)23 (82.1%)14 (70.0%)18 (72.0%)0.0536 (60.0%)5 (20.0%)0.0216.0001.211–29.7408 (47.1%)3 (16.7%)0.0534 (40.0%)20 (80.0%)9 (52.9%)15 (83.3%)9 (52.9%)15 (83.3%)0.0537 (58.3%)3 (14.3%)0.0088.4001.570–44.9004 (21.1%)6 (42.9%)0.1805 (41.7%)18 (85.7%)15 (78.9%)8 (57.1%)8 (57.1%)0.0106.6671.476–30.1207 (38.9%)5 (25.0%)0.3586 (42.9%)20 (83.3%)11 (61.1%)15 (75.0%)15 (75.0%)0.103	GGGA+AAp-valueOR95%CI $CT+TT$ CCp-valueOR8 (47.1%)5 (17.9%)0.0364.0891.052-15.8906 (30.0%)7 (28.0%)0.8831.1029 (52.9%)23 (82.1%)14 (70.0%)18 (72.0%)18 (72.0%)0.0534.4406 (60.0%)5 (20.0%)0.0216.0001.211-29.7408 (47.1%)3 (16.7%)0.0534.4404 (40.0%)20 (80.0%)9 (52.9%)15 (83.3%)15 (83.3%)0.0534.4407 (58.3%)3 (14.3%)0.0088.4001.570-44.9004 (21.1%)6 (42.9%)0.1800.3565 (41.7%)18 (85.7%)15 (78.9%)8 (57.1%)8 (57.1%)0.0106.6671.476-30.1207 (38.9%)5 (25.0%)0.3581.9096 (42.9%)20 (83.3%)0.0146.6001.339-32.5304 (19.0%)7 (43.8%)0.1030.302

## Discussion

This study explored the correlation of *USF1* gene polymorphisms with the efficacy and safety of paclitaxelbased chemotherapy and prognosis in the treatment of OC, and the results indicate that the rs2516839 polymorphisms in USF1 gene might have an association with the increase of paclitaxel resistance of OC, which was firstly proposed. The Logistic regression analysis has presented that there was a significant difference in the genotype and allele frequencies of rs2516839 between the sensitive and resistant groups and no obvious difference in the genotypes and allele frequencies of rs3737787 was found between these two groups. In the previous study, it has been demonstrated that *TEKT4* 

Table 6. Relationship between the USF1 gene polymorphism and the prognosis of OC patients

The of Readonship between the cost i gene polymorphism and the prognosis of cost patients.									
Variable	Regression coefficient (B)	Standard error	p-value	OR	95%CI				
rs2516839(G>A)	0.975	0.302	0.001	2.652	1.466-4.797				
Pathology	0.275	0.111	0.013	1.317	1.060-1.635				
Physical status score	0.756	0.290	0.009	2.129	1.205-3.761				
Maximum tumor diameter	0.442	0.152	0.004	1.555	1.156-2.094				

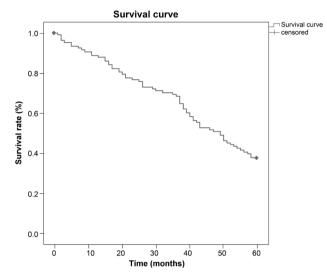


Figure 1. Kaplan-Meier curves of cumulative survival rates of 100 OC patients.

gene could cause a poorer response and survival of patients treated with paclitaxel-based neoadjuvant chemotherapy [10]. It has also been reported that the rs2516838 polymorphisms dominant model and allelic model were protective factors in papillary thyroid cancer [17]. And other results indicated that the SNP of rs2516839 was closely correlated with the increased risk of hepatocellular carcinoma [18].

According to Cox multiple regression analysis, our study demonstrates that USF1 gene affects the prognosis of OC patients and rs2516839 in USF1 gene is one of the independent risk factors for the prognosis of OC patients. Physical status score, tumor pathology, and maximum tumor diameter were also independent risk factors for the prognosis of OC patients. And physical status score (2 points), maximum tumor diameter (larger than 2 cm), serous adenocarcinoma and GA+AA carriers at rs2516839 were risk factors for the prognosis of OC patients. This was the first study about the association between USF1 and prognosis of OC patients. Former study had suggested that stage at diagnosis, maximum residual disease following cytoreductive surgery, and performance status were the three major prognostic factors of OC [19]. Besides, it has been defined that the expression of G protein-coupled receptor 56, size of residual tumor at the end of primary

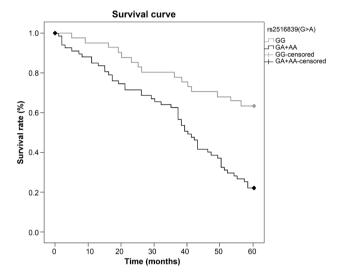


Figure 2. Kaplan-Meier curves of OC patients carrying different genotypes of rs2516839 in USF1 gene.

surgery and age at diagnosis were independent prognostic factors in OC [20, 21]. Moreover, it also has been reported that age, performance status, tumor histology, optimal cytoreduction, and chemotherapy were key prognostic factors for OC patients. [4].

With regard to the safety of paclitaxel-based chemotherapy, no toxicity-related death was found. There was no significant difference in toxic reaction of OC patients between the sensitive and resistant groups. Furthermore, stratified analysis to 100 OC patients showed that the incidences of serious toxic reactions in patients carrying GA+AA genotype of rs2516839 in USF1 gene were significantly higher than in the patients carrying GG genotype (p<0.05). It has been reported that no cancer-free patient has developed late bowel toxicity >CTC 2 in the study about carboplatin-paclitaxel based chemotherapy for advanced uterine epithelial cancer, and all patients treated with radiotherapy showed no toxicity-related interruptions and no CTCAE grade 4 toxicities occurred during the carboplatinpaclitaxel based chemotherapy [22]. A study has demonstrated that the albumin-bound paclitaxel-based chemotherapy is safe and efficient in the operation for patients with advanced esophageal cancer and no unanticipated treatment or toxicity-related deaths were observed [23]. What's more,

there emerged a variety of new drugs on the safety and efficacy of the treatment of cerebral ischemia and cancer has sought more and further investigations in future study, such as 3-n-Butylphthalide (NBP) and Famitinib, which were well absorbed and extensively metabolized by multiple enzymes to various metabolites prior to urinary excretions [24, 25].

In summary, our preliminary findings suggest that the rs2516839 polymorphism in USF1 gene might associate with the efficacy and safety of paclitaxel-based chemotherapy and prognosis in treating OC, which provides a new idea and new way in the operation of OC patients. However, there were a few limitations in the present study such as a trial size in the designed experiment, which was relatively small and gene-gene or gene-environment interactions which may influence the results. It must be noted that our findings need to be further validated by more specific studies with larger samples, more advanced gene technologies and well-designed chemotherapy. We will extend the time period of our study to involve more subjects and further study the correlation of rs2516839 polymorphism in USF1 gene with the efficacy and safety of paclitaxel-based chemotherapy and prognosis in the treatment of OC.

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