

CircABC10 silencing inhibits the cell ferroptosis and apoptosis by regulating the miR-326/CCL5 axis in rectal cancer

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Circular ATP binding cassette subfamily B member 10 (circABC10) has been identified to have oncological functions in several tumors. However, the roles of circABC10 in rectal cancer remain unknown. The expression of circABC10, microRNA (miR)-326 and C-C motif chemokine ligand 5 (CCL5), and apoptosis related-protein was detected using quantitative real-time polymerase chain reaction or western blot, respectively. Cell survival or apoptosis was measured using cell counting kit-8 assay or flow cytometry. The accumulations of intracellular lipid reactive oxygen species (ROS) and Fe²⁺ were analyzed using C11-BODIPY dye or iron kit assay, respectively. *In vivo* experiments were conducted using the murine xenograft model. The interaction between miR-326 and circABC10 or CCL5 was confirmed by dual-luciferase reporter assay and RNA immunoprecipitation assay. CircABC10 and CCL5 were upregulated but miR-326 was downregulated in rectal cancer. The knockdown of circABC10 promoted rectal cancer cell ferroptosis and apoptosis *in vitro* as well as inhibited tumor growth *in vivo*. miR-326 was a target of circABC10, and the miR-326 inhibition could partially attenuate circABC10 deletion-induced cell ferroptosis and apoptosis. miR-326 directly interacted with CCL5, and the miR-326 inhibition suppressed cell ferroptosis and apoptosis by targeting CCL5. Besides, we observed that miR-326 was negatively regulated by circABC10, while CCL5 was positively regulated by it, and circABC10 served as a sponge of miR-326 to regulate the CCL5 expression in rectal cancer cells. CircABC10 silencing promoted rectal cancer cell ferroptosis and apoptosis by regulating the miR-326/CCL5 axis, suggesting a potential therapeutic target for rectal cancer therapy.

Key words: circABC10, miR-326, CCL5, apoptosis, ferroptosis, rectal cancer

Colorectal cancer (CRC) is the third most common solid malignancy and the fourth highest leading cause of cancer-related mortality worldwide [1]. Rectal cancer is one of the main sub-types of CRC, accounting for around 30% of CRC, with a high risk of metastasis and relapse [2, 3]. With the advances in the multimodality treatment of rectal cancer, locally advanced rectal cancer patients have better survival rates than patients with colon cancer, but at the expense of munificent morbidities and lower quality of life [4]. Thus, further investigations on the understanding of the molecular mechanisms involved in the pathogenesis of rectal cancer are necessary to develop ideal diagnostic or therapeutic biomarkers for early detection and treatment of rectal cancer.

Increasing evidence reports that diverse types of RNAs have emerged as important regulators in the development and progression of tumors by acting in multiple ways [5, 6]. Circular RNAs (circRNAs) are one type of highly conserved RNA transcripts with covalently closed loop structures

and are formed by the back-splicing of precursor mRNA (pre-mRNA), which make them resistant to regular mechanisms of linear RNAs decay [7, 8]. Recent studies have shown that circRNAs are dysfunctional in diverse human cancers and dysregulated circRNAs play important roles in the tumorigenesis and tumor progression in a variety of cancers, including CRC [9–11]. Circ ATP binding cassette subfamily B member 10 (circABC10) is a novel-identified circRNA, which has been indicated to have oncological functions to promote cell proliferation and progression in breast cancer, clear cell renal cell carcinoma, and epithelial ovarian cancer [12–14]. However, the roles of circABC10 in rectal cancer remain largely unknown.

MicroRNAs (miRNAs) are the best-characterized type of RNAs, which can interact with the complementary sequences in the 3'-untranslated region (UTR) of their target mRNAs to post-transcriptionally control gene expression [15]. It has been demonstrated that circRNAs function as

sponges of miRNA, affect the expression of miRNAs and the competition between mRNAs [8, 16], thereby involving in the initiation and progression of a variety of cancers [11, 17, 18]. miR-326 is a well-recognized tumor suppressor, which has been found to be downregulated and associated with the tumorigenesis of CRC and is a potential therapeutic target for CRC [19]. C-C motif chemokine ligand 5 (CCL5) is known as a chemotactic cytokine, whose expression and secretion are regulated in T cells [20]. Although CCL5 has been initially identified as an inducer to recruit leukocytes to inflammatory sites [21], increasing findings documented the high expression of CCL5 in a variety of cancers, and abnormal CCL5 expression contributed to promote tumor development and metastasis in many cancers, including CRC [22, 23]. However, the exact molecular mechanism of CCL5 in rectal cancer is still unclear.

Cell death is an important biological process, which can maintain homeostasis during development and disease prevention [24]. Ferroptosis and apoptosis are two modes of cell death and have been identified to play important roles in the tumorigenesis of cancers [25, 26]. Therefore, this study focused on the expression and functions of circABC10, miR-326, and CCL5 in the ferroptosis and apoptosis of rectal cancer, and explored the potential regulatory relationship among them in cell ferroptosis and apoptosis in rectal cancer.

Materials and methods

Clinical samples. This study was approved by the Ethics Committee of The Sixth Affiliated Hospital of Sun Yat-sen University. All patients had signed informed consent before the study. Tumor tissues and adjacent non-tumor tissues from 30 rectal cancer patients who underwent surgical resection were obtained from The Sixth Affiliated Hospital of Sun Yat-sen University and were immediately stored at -80°C for further analysis. All patients were diagnosed by histopathological examination and did not receive any preoperative treatment.

Cell culture. Human rectal mucosa epithelium cell line (FHC) and rectal cancer cell lines SW837 and HR-8348 were obtained from the Shanghai Academy of Life Science (Shanghai, China) and maintained in the Roswell Park Memorial Institute-1640 (RPMI-1640, Gibco, Carlsbad, CA, USA) medium supplemented with 10% fetal bovine serum (FBS, Gibco), 100 U/ml penicillin and 100 mg/ml streptomycin at 37°C with 5% CO_2 .

Cell transfection. The short hairpin RNA (shRNA) targeting circABC10 covalent closed junction (sh-circABC10), shRNA scramble control (sh-NC), the small interfering RNA (siRNA) sequences targeting circABC10 covalent closed junction (si-circABC10, 5'-TAGAAGAC-CATGGGGGATGTCAAGAGCATCCCCATGGTCTTC-TATTTTTT-3'), siRNA sequences targeting CCL5 (si-CCL5, 5'-UCUUUCUGGUGAUAAGACGAGUCUUUAUCAC-

CAGAAAGAAC-3'), siRNA negative control (si-NC) were designed by Genepharma (Shanghai, China). The miR-326 mimic (miR-326), miR-326 inhibitor (anti-miR-326), and their corresponding negative control (miR-NC and anti-miR-NC) were purchased from RIBOBIO (Guangzhou, China). The transfection of plasmids or miRNAs was carried out using Lipofectamine™ 2000 transfection reagent (Invitrogen, Carlsbad, CA, USA).

Quantitative real-time polymerase chain reaction (qRT-PCR). Trizol reagent (Invitrogen) was used to extract total RNA according to the standard procedure. Subsequently, isolated RNA was incubated with Rnase R (Epicentre, Madison, WI, USA), followed by the interaction with RNeasy Min Elute Cleanup Kit (Qiagen, Valencia, CA, USA). Complementary DNA (cDNA) was synthesized using the Prime Script RT Master Mix (Applied Biosystems, Foster City, CA, USA), and then quantitative PCR was carried out with SYBR Premix Ex Taq (Qiagen). Relative transcription alterations were analyzed by $2^{-\Delta\Delta\text{Ct}}$ method and normalized by glyceraldehyde 3-phosphate dehydrogenase (GADPH) or U6 small nuclear B noncoding RNA (U6). The specific primer sequences were presented as follow: circ-ABC10: F 5'-CTAAGGAGTCACAGGAAGACATC-3', R 5'-GTAGAATCTCTCAGACTCAAGGTTG-3'; CCL5: F 5'-CCATGAAGGTCTCCGCGGCAC-3', R 5'-CCTAGCT-CATCTCCAAAGAG-3'; miR-326: F 5'-CCTCTGGGCC-CTTCCTCCAG-3', R 5'-GCTGTCAACGATACGCTAC-CTA-3'; GADPH: F 5'-GATATGTTGCCATCAATGAC-3', R 5'-TTGATTTTGGAGGGATCTCG-3'; U6: F 5'-CTCGCT-TCGGCAGCACA-3', R 5'-ACGCTTCACGAATTTGCGT-3'.

Cell viability assay. Transfected SW837 and HR-8348 cells were seeded into the wells of a 96-well plate. After stimulation with erastin (Selleck Chemicals, Houston, TX, USA; 0–40 μM) for 24 h, 10 μl of cell counting kit-8 (CKK-8) solution (Beyotime, Shanghai, China) was added into each well and incubated for another 2 h. Finally, the reading of the optical density (OD) was detected on a microplate reader at 450 nm to assess the survival rate.

Lipid reactive oxygen species (ROS) measurement. Transfected cells were treated with 5 μM erastin for 24 h, and then were resuspended in medium with 10% FBS (Gibco). Immediately, 10 μM C11-BODIPY (Thermo Fisher Scientific, Inc., Waltham, MA, USA) was added to the cell suspension and incubated for 30 min at 37°C in the dark. After washing with PBS twice, the fluorescence intensity of the samples was measured by using a flow cytometer.

Iron assay. The level of intracellular Fe^{2+} was detected by the iron assay kit (Abcam, Cambridge, MA, USA) following the standard procedure. In brief, transfected cells were treated with 5 μM erastin for 24 h. Cells were collected and washed with cold PBS and then homogenized in 4–10 volumes of the iron assay buffer. After removing the insoluble material, 5 μl iron reducer was added to each well before mixing, followed by incubation for 30 min at 37°C in the dark. Subsequently, each well was added with 100 μl of the iron probe and mixed,

and then incubated for 60 min at 37°C in dark conditions. The OD was detected at 593 nm using a colorimetric microplate reader. Besides, total protein concentration in 50 µl samples was quantified using the bicinchoninic acid assay, and the level of Fe²⁺ was assessed with the following equation: level = volume (nmol/µl)/concentration (mg/µl).

Cell apoptosis assay. Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) apoptosis detection kit (BD Biosciences, San Jose, CA, USA) was used to detect apoptotic cells following the standard protocol. Transfected cells were resuspended in binding buffer, followed by staining with 5 µl FITC annexin V and 10 µl PI. Finally, the apoptotic cells were measured by FlowJo software.

Western blot analysis. Transfected cells were collected and then lysed using the radio-immunoprecipitation assay (RIPA) buffer (Beyotime). Cell lysates were separated on 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis, transferred onto a polyvinylidene fluoride membrane, and blocked with 5% non-milk. Subsequently, the membranes were incubated with primary antibodies against B-cell lymphoma-2 (Bcl-2, 1:500, ab692, Abcam), Bcl2-associated X protein (Bax, 1:1000, ab32503, Abcam), CCL5 (1:1000, ab9679, Abcam) and β-Actin (1:1000, 4967, Cell Signaling Technology, Boston, MA, USA), followed by incubation with secondary HRP-conjugated antibody (1:1000, ab9482, Abcam). Finally, signals were visualized using the chemiluminescence chromogenic substrate (Beyotime).

In vivo experiments. BALB/c nude mice (female, 4–6 weeks of age, N=6) were used to perform the xenograft assays. HR-8348 cells were stably transfected with lentivirus containing sh-circ-ABC10 or sh-NC and then subcutaneously injected into the flanks of the nude mice. After one week following the inoculation, the tumor size was examined every week and the tumor volume was calculated. After 4 weeks, all mice were sacrificed and tumor masses were weighted and harvested for further molecular analysis. This study was approved by the Animal Research Committee of The Sixth Affiliated Hospital of Sun Yat-sen University and undertaken in accordance with the guidelines of the National Animal Care and Ethics Institution.

Dual-luciferase reporter assay. The wild-type (WT) or mutant (MUT) circ-ABC10/CCL5 3'UTR possessing miR-326 binding sequences were amplified and cloned into the pmirGLO Vector (Promega, Shanghai, China). Then, SW837 and HR-8348 cells were co-transfected with the constructed vectors and miR-326 or miR-NC using Lipofectamine 2000 reagent (Invitrogen) and incubated for 48 h. Finally, a dual-luciferase reporter assay kit (Promega) was used to measure the relative luciferase activity.

RNA immunoprecipitation (RIP) assay. RIP assay was performed with the Magna RIP Kit (Millipore, Billerica, MA, USA). SW837 and HR-8348 cells transfected with miR-326 or miR-NC were pelleted and lysed in RIP buffer (Millipore), and then the lysate was incubated with magnetic beads coated with human anti-Ago2 or IgG antibody. Subsequently,

the enrichment of circ-ABC10 or CCL5 was measured by qRT-PCR as described above, respectively.

Statistical analysis. All statistical data from triplicate repeat independent experiment were expressed as a mean ± standard deviation (SD) and analyzed using GraphPad Prism 7 software (GraphPad Inc., San Diego, CA, USA). The differences in different groups were analyzed by Student's t-test or one-way analysis of variance (ANOVA). The correlation analysis among circ-ABC10, miR-326, and CCL5 was performed using Spearman's rank correlation. A p-value <0.05 presented statistically significant.

Results

CircABC10 is upregulated in rectal cancer and its depletion regulates erastin-induced ferroptosis in rectal cancer. To explore the roles of circABC10 in rectal cancer, we firstly detected the expression of circABC10 in 30 paired rectal cancer patients, and results showed circABC10 was higher in rectal cancer tissues compared to the normal tissues (Figure 1A). Similarly, the qRT-PCR analysis also indicated an upregulation of circABC10 in rectal cancer cell lines SW837 and HR-8348 compared with the rectal mucosa epithelium cell line (FHC) (Figure 1B). Erastin is a classical small-molecule inducer of ferroptosis [27]. To detect whether circABC10 could regulate ferroptosis, circABC10 was inhibited in SW837 and HR-8348 cells using siRNA sequences. As expected, circABC10 was significantly reduced in SW837 and HR-8348 cells (Figure 1C). After treatment with erastin (0–40 µM) for 24 h, CCK-8 assay indicated erastin-induced cell death in SW837 and HR-8348 cells in a dose-dependent manner and circABC10 depletion significantly increased erastin-induced ferroptosis both in SW837 and HR-8348 cells (Figures 1D, 1E), suggesting that circABC10 might be an important regulator of ferroptosis in rectal cancer cells.

CircABC10 depletion promotes rectal cancer cell ferroptosis and apoptosis *in vitro* as well as inhibits tumor growth *in vivo*. Ferroptosis is involved in the generation of iron-dependent ROS, thus we investigated the accumulation of lipid ROS and Fe²⁺, which are two essential factors in triggering ferroptosis [27], in rectal cancer cells treated with 5 µM erastin for 24 h. We found that circABC10 depletion significantly promoted the accumulation of intracellular lipid ROS in SW837 and HR-8348 cells (Figure 2A). Furthermore, the knockdown of circABC10 also increased the intracellular concentrations of Fe²⁺ in SW837 and HR-8348 cells (Figure 2B), indicating circABC10 depletion induced sensitivity to ferroptosis in rectal cancer cells. Besides that, we observed that circABC10 depletion induced cell apoptosis in SW837 and HR-8348 cells (Figure 2C). Moreover, western blot showed circABC10 silence increased the level of Bax, but decreased the level of Bcl-2 in SW837 and HR-8348 cells (Figure 2D), further implying circABC10 silence induced cell apoptosis in rectal cancer.

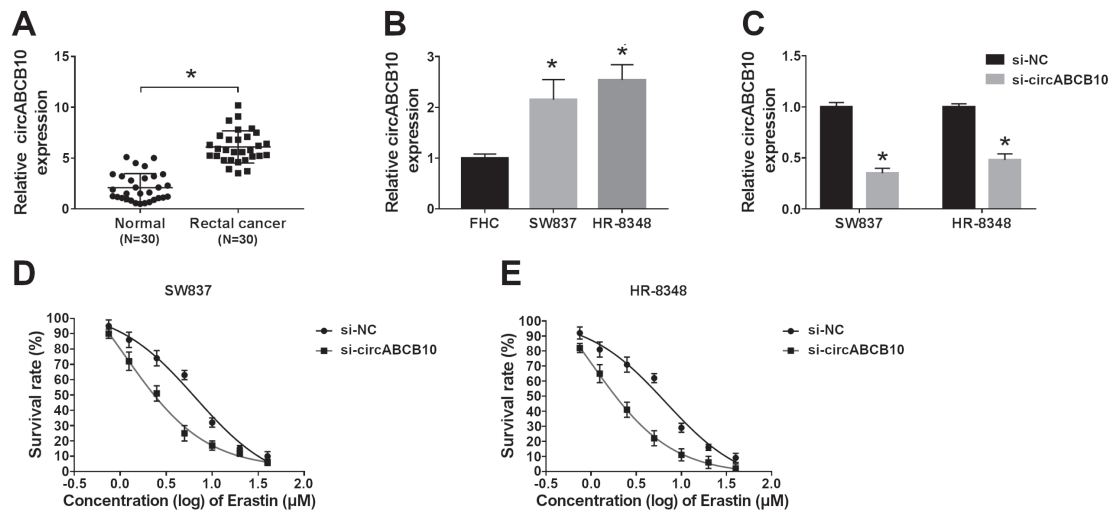


Figure 1. CircABC10 is upregulated in rectal cancer and circABC10 depletion regulates erastin-induced ferroptosis in rectal cancer. A, B) The expression of circABC10 was detected using qRT-PCR in rectal cancer tumor tissues, normal tissues, rectal cancer cell lines SW837 and HR-8348, as well as rectal mucosa epithelium cell line (FHC; Student's t test). circABC10 was inhibited in SW837 and HR-8348 cells using siRNA sequences. C) The interference efficiency was measured using qRT-PCR (Student's t test). D, E) SW837 and HR-8348 cells were treated with erastin (0–40 μ M) for 24 h and cell death was assayed using a CCK-8 kit (Student's t test). * $p < 0.05$

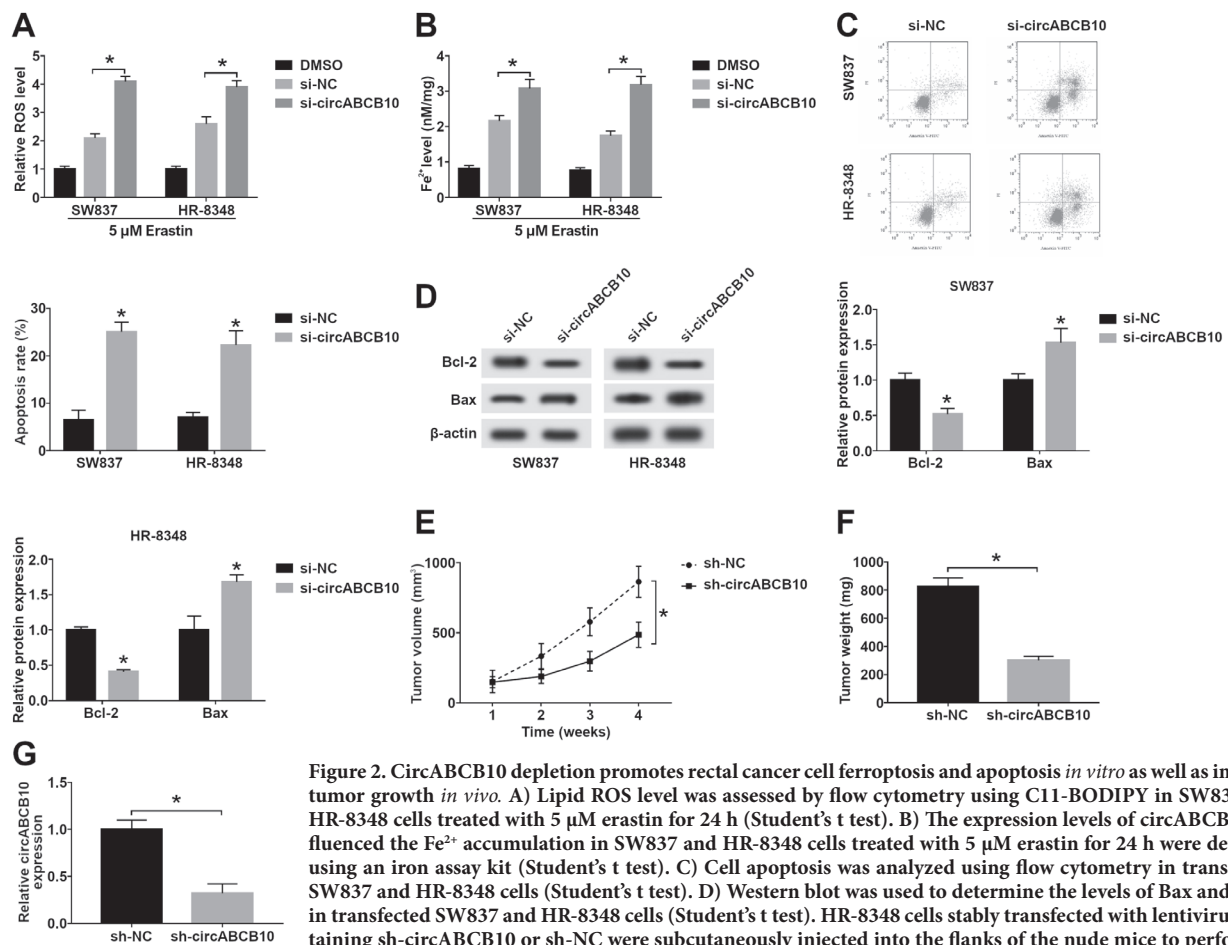


Figure 2. CircABC10 depletion promotes rectal cancer cell ferroptosis and apoptosis *in vitro* as well as inhibits tumor growth *in vivo*. A) Lipid ROS level was assessed by flow cytometry using C11-BODIPY in SW837 and HR-8348 cells treated with 5 μ M erastin for 24 h (Student's t test). B) The expression levels of circABC10 influenced the Fe^{2+} accumulation in SW837 and HR-8348 cells treated with 5 μ M erastin for 24 h were detected using an iron assay kit (Student's t test). C) Cell apoptosis was analyzed using flow cytometry in transfected SW837 and HR-8348 cells (Student's t test). D) Western blot was used to determine the levels of Bax and Bcl-2 in transfected SW837 and HR-8348 cells (Student's t test). HR-8348 cells stably transfected with lentivirus containing sh-circABC10 or sh-NC were subcutaneously injected into the flanks of the nude mice to perform *in vivo* experiments. E) Tumor volumes were calculated every week (Student's t test). F) Mice were euthanized on day 28 after infection, and then tumor masses were excised and weighed (Student's t test). G) The expression of circABC10 in excised tumor masses was determined using qRT-PCR (Student's t test). * $p < 0.05$

After that, HR-8348 cells stably transfected with lenti-virus containing sh-circABC10 or sh-NC were subcutaneously injected into the flanks of the nude mice to perform *in vivo* experiments. We observed that circABC10 depletion markedly impaired tumor volume, weight, and inhibited circABC10 expression in the sh-circABC10 group (Figures 2E–2G). Taken together, our results demonstrated that circABC10 was associated with cell apoptosis, ferroptosis, and tumor growth *in vivo* and *in vitro*, suggesting an oncogenic role of circABC10 in rectal cancer progression.

CircABC10 is a sponge of miR-326 and regulates miR-326 expression. To detect the potential pathways by which circABC10 regulated cell ferroptosis and apoptosis in rectal cancer, the potential miRNA targets of circABC10 were searched using the bioinformatics tool starBase program and miR-326 was predicted to contain putative binding sites of circABC10 (Figure 3A). To verify the bioinformatics-based prediction, a dual-luciferase activity assay was conducted and results showed miR-326 overexpression dramatically reduced the luciferase activity of WT circABC10 reporter vector, but not MUT circABC10 reporter vector in SW837 and HR-8348 cells (Figure 3B). Furthermore, RIP assay using anti-Ago2 antibody also confirmed that miR-326 interacted with circABC10 in SW837 and HR-8348 cells due to the enrichment of circABC10 (Figure 3C). In addition,

qRT-PCR exhibited that the knockdown of circABC10 promoted miR-326 expression in SW837 and HR-8348 cells (Figure 3D). These data suggested circABC10 as a sponge of miR-326 in rectal cancer cells. In the meanwhile, miR-326 was found to be decreased in rectal cancer tissues and cell lines (Figures 3E, 3F), and a negative correlation between miR-326 and circABC10 was observed in tumor tissues (Figure 3G), suggesting miR-326 might be involved in circABC10-induced regulation on rectal cancer cells.

CircABC10 depletion promotes rectal cancer cell ferroptosis and apoptosis by interacting with miR-326. Based on the relationship between circABC10 and miR-326, we explored whether the circABC10/miR-326 axis was responsible for rectal cancer cell ferroptosis and apoptosis. First, SW837 and HR-8348 cells were transfected with the anti-miR-326 and anti-miR-NC, and a significantly decreased miR-326 was found in SW837 and HR-8348 cells (Figure 4A). Next, SW837 and HR-8348 cells were transfected with anti-miR-NC, anti-miR-326, si-circABC10+anti-miR-NC, or si-circABC10+anti-miR-326 to conduct the rescue assay. Subsequently, results indicated miR-326 inhibition significantly decreased erastin-induced ferroptosis (Figure 4B), and the accumulation of intracellular lipid ROS (Figure 4C) as well as Fe²⁺ (Figure 4D) both in SW837 and HR-8348 cells. Furthermore, miR-326 inhibition also suppressed

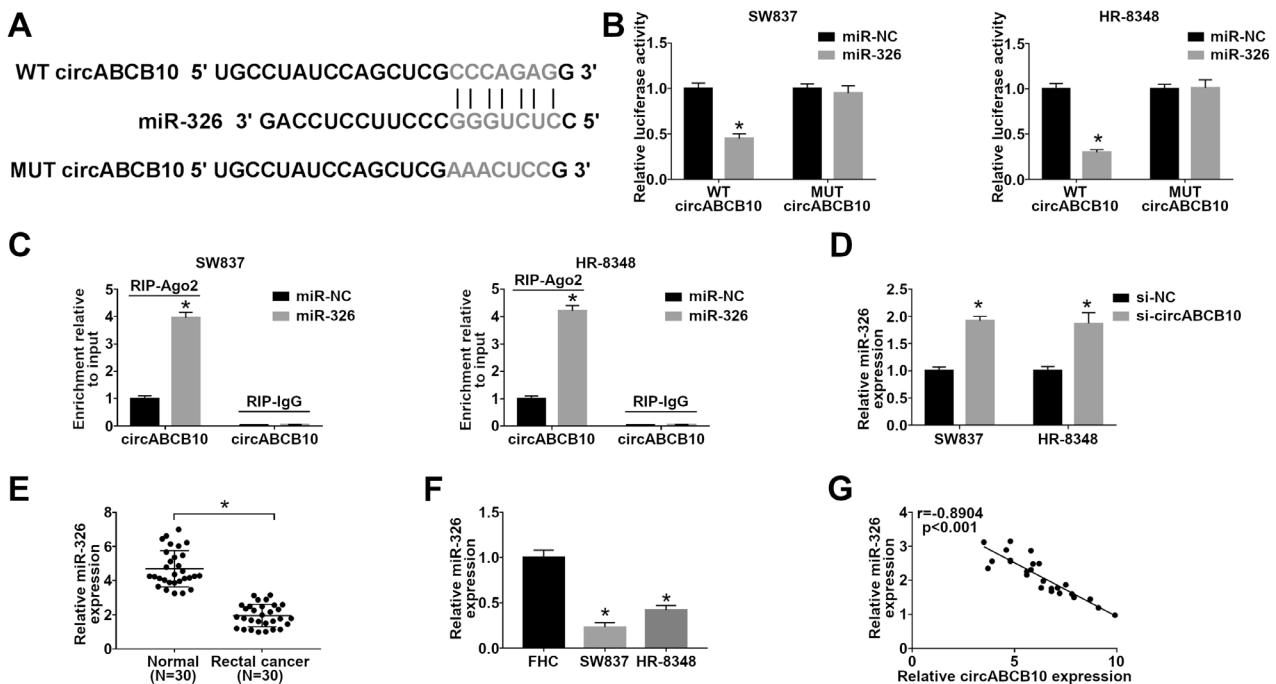


Figure 3. circABC10 is a sponge of miR-326 and regulates miR-326 expression. A) The putative binding sites between circABC10 and miR-326 were listed. B) Luciferase activity was measured in SW837 and HR-8348 cells co-transfected with WT circABC10 or MUT circABC10 and miR-NC or miR-326 (Student's t test). C) The enrichment of circABC10 was detected in SW837 and HR-8348 cells transfected with miR-NC or miR-326 after RIP (Student's t test). D) The level of miR-326 was assessed using qRT-PCR in SW837 and HR-8348 cells transfected with si-NC or si-circABC10 (Student's t test). E, F) qRT-PCR was applied to measure the level of miR-326 in rectal cancer tumor tissues and cell lines (Student's t test). G) The correlation between circABC10 and miR-326 was analyzed using Spearman's rank correlation. * $p < 0.05$

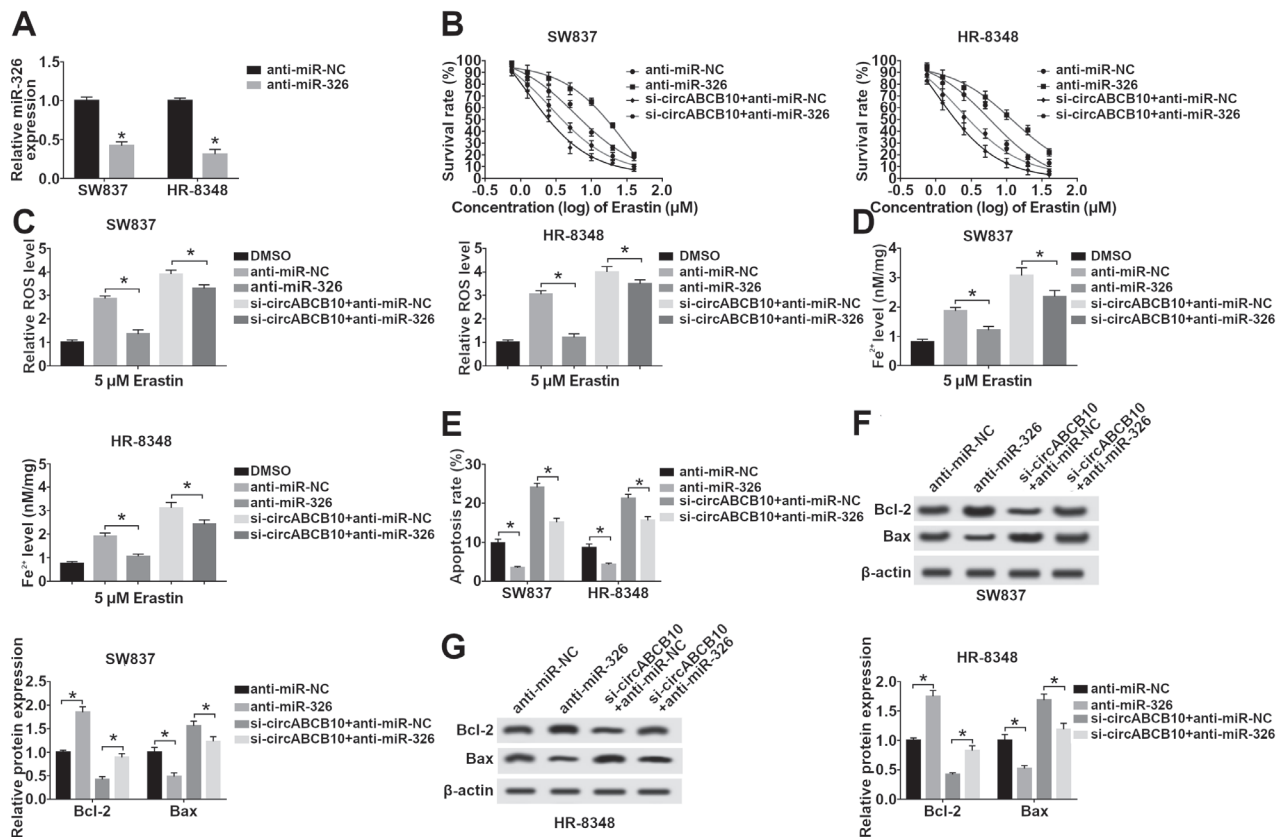


Figure 4. circABC10 depletion promotes rectal cancer cell ferroptosis and apoptosis by interacting with miR-326. SW837 and HR-8348 cells were transfected with anti-miR-NC or anti-miR-326. A) The interference efficiency was measured using qRT-PCR (Student's t test). B) Cell death was assayed using a CCK-8 kit in SW837 and HR-8348 cells treated with erastin (0–40 μ M) for 24 h (One-way ANOVA). C) Lipid ROS level was assessed by flow cytometry using C11-BODIPY in SW837 and HR-8348 cells treated with 5 μ M erastin for 24 h (One-way ANOVA). D) The expression levels of the Fe^{2+} accumulation in SW837 and HR-8348 cells treated with 5 μ M erastin for 24 h were detected using an iron assay kit (One-way ANOVA). E) Cell apoptosis was analyzed using flow cytometry in transfected SW837 and HR-8348 cells (One-way ANOVA). F, G) Western blot was used to determine the level of Bax and Bcl-2 in transfected SW837 and HR-8348 cells (One-way ANOVA). * $p < 0.05$

SW837 and HR-8348 cell apoptosis, demonstrated by the decreased apoptotic cells (Figure 4E) and Bax expression (Figures 4F, 4G), as well as the increased expression of Bcl-2 (Figures 4F, 4G). Meanwhile, the rescue assay showed miR-326 inhibition could partially reverse circABC10 depletion-induced promotion on cell ferroptosis (Figures 4B–4D) and apoptosis (Figures 4E–4G) in SW837 and HR-8348 cells. These results suggested circABC10 depletion promoted rectal cancer cell ferroptosis and apoptosis by serving as a sponge of miR-326.

CCL5 is a target of miR-326 and circABC10 indirectly regulates CCL5 by binding to miR-326. We further elucidated the mechanisms underlying the circABC10/miR-326 mediated ferroptosis and apoptosis of rectal cancer cells, thus the target genes of miR-326 were searched. Using the bioinformatics tool TargetScan program, we found CCL5 contained the putative binding sequences of miR-326 (Figure 5A). To confirm the interaction, we performed a dual-luciferase reporter assay and found the luciferase

activity of the WT-CCL5 reporter was obviously repressed, but there was no notable change in MUT-CCL5 reporter after overexpression of miR-326 in SW837 and HR-8348 cells (Figure 5B). In the meanwhile, using a RIP assay, we evaluated the CCL5 level in SW837 and HR-8348 cells after miR-326 immunoprecipitation using IgG as controls and found CCL5 was enriched after miR-326 immunoprecipitation in SW837 and HR-8348 cells, further suggesting the interaction between miR-326 and CCL5 (Figure 5C). Besides that, we demonstrated circABC10 depletion inhibited CCL5 expression using qRT-PCR and western blot assay, while this inhibition could be rescued by the inhibition of miR-326 in SW837 and HR-8348 cells (Figures 5D, 5E). Therefore, a circABC10/miR-326/CCL5 axis was identified in rectal cancer cells.

miR-326 inhibition inhibits cell ferroptosis and apoptosis by interacting with CCL5 in rectal cancer. Given that miR-326 targeted CCL5, we hypothesized that CCL5 might influence miR-326 induced regulation on cell

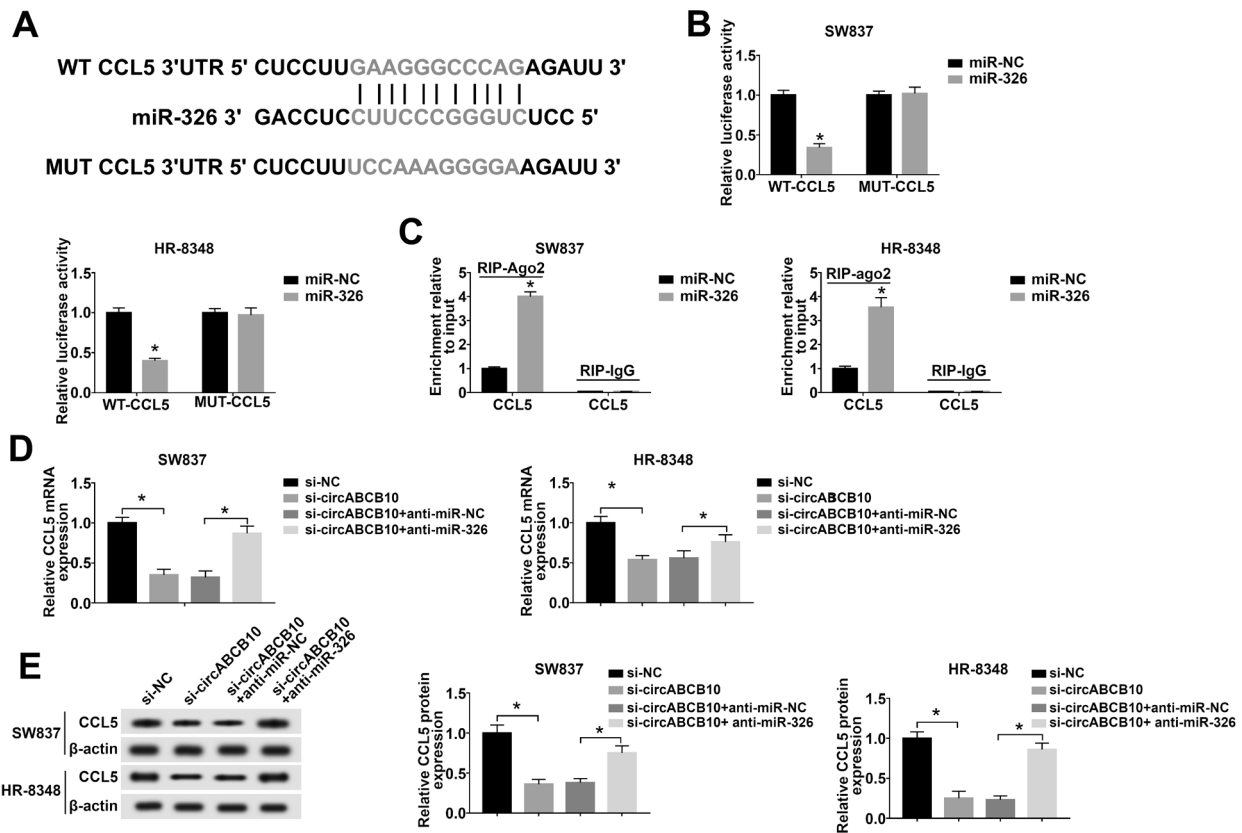


Figure 5. circABC10 indirectly regulates CCL5 by binding to miR-326. **A**) The putative binding sites between CCL5 and miR-326 were presented. **B**) Luciferase activity was measured in SW837 and HR-8348 cells co-transfected with WT-CCL5 or MUT-CCL5 and miR-NC or miR-326 (Student's *t* test). **C**) The enrichment of CCL5 was detected in SW837 and HR-8348 cells transfected with miR-NC or miR-326 after RIP (Student's *t* test). SW837 and HR-8348 cells were transfected with si-NC, si-circABC10, si-circABC10+anti-miR-NC, or si-circABC10+anti-miR-326. **D, E**) The level of CCL5 at mRNA and protein level was determined using qRT-PCR and western blot assay in SW837 and HR-8348 cells after treatment (One-way ANOVA). **p*<0.05

ferroptosis and apoptosis in rectal cancer. First, we determined CCL5 expression at mRNA and protein levels, and results showed CCL5 was high in rectal cancer tumor tissues and cell lines compared with the controls (Figures 6A–6D). Moreover, a negative correlation between CCL5 and miR-326 (Figure 6E), and a positive correlation between CCL5 and circABC10 (Figure 6F) were observed, indicating the regulatory relationship among the circABC10, miR-326, and CCL5. Subsequently, CCL5 was suppressed in SW837 and HR-8348 cells using siRNA sequences, and CCL5 expectedly decreased in SW837 and HR-8348 cells at mRNA and protein levels (Figures 6G, 6H).

After that, functional experiments were performed. We found CCL5 silence increased erastin-induced ferroptosis (Figure 7A), and the intracellular accumulation of intracellular lipid ROS (Figure 7B) and Fe²⁺ (Figure 7C) both in SW837 and HR-8348 cells. Additionally, CCL5 silence also promoted cell apoptosis of SW837 and HR-8348 cells (Figure 7D), and western blot analysis indicated CCL5

silence reduced the level of Bcl-2, but enhanced the level of Bax in SW837 and HR-8348 cells (Figures 7E, 7F), further demonstrating CCL5 silence induced apoptosis of SW837 and HR-8348 cells. However, the rescue assay showed CCL5 silence could partially overturn miR-326 inhibition-induced repression on cell ferroptosis (Figures 7A–7C) and apoptosis (Figures 7D–7F) in SW837 and HR-8348 cells. Altogether, miR-326 regulated cell ferroptosis and apoptosis by interacting with CCL5 in rectal cancer.

Discussion

Recently, with the development of high-throughput technologies, emerging evidence indicates the pivotal role of circRNAs in the tumorigenesis of cancers [28]. Until now, many circRNAs have been identified to be involved in the development and progression of CRC. For example, circ_0026344 suppressed cell migration, invasion, and epithelial-mesenchymal transition (EMT) process by directly

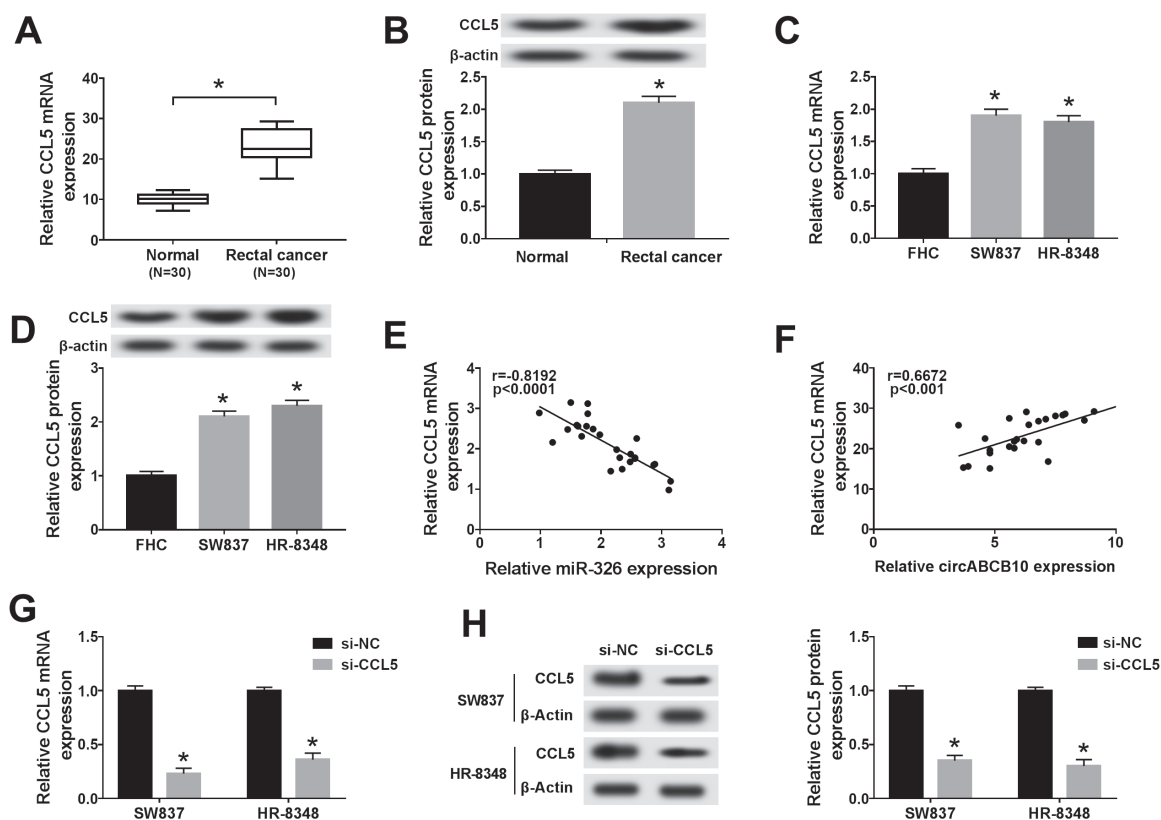


Figure 6. CCL5 is upregulated in rectal cancer. (A–D) The expression of CCL5 was detected in rectal cancer tumor tissues and cell lines at mRNA and protein levels using qRT-PCR or western blot assay (Student's t test). (E, F) The correlation between CCL5 and miR-326 or circABC10 was assessed using Spearman's rank correlation. (G, H) The expression of CCL5 was measured in SW837 and HR-8348 cells transfected with si-CCL5 or si-NC using qRT-PCR and western blot assay (Student's t test). * $p < 0.05$.

interacting with miR-183 in CRC [29]. Circ_0000218 performed carcinogenic effects to regulate cell proliferation and metastasis in CRC through the miR-139-3p/RAB1A axis [30]. Circ_0021977 served as a tumor suppressor to inhibit cell metastasis and proliferation in CRC by regulating the expression of p21 and p53 through miR-10b-5p [31]. Therefore, circRNAs may be promising candidates for the progression and therapy of CRC.

Cell death is a primary element of cellular metabolism and is necessary for the development and prevention of disease [32]. Previous evidence documented that apoptosis was the only regulated form of cell death [25], but recent studies have indicated several non-apoptotic cell death pathways, such as ferroptosis [27]. Ferroptosis is an oxidative, iron-dependent mode of non-apoptotic cell death, which induces the inactivation of glutathione and the production of toxic lipid ROS. Herein, we explored circABC10-related epigenetic regulation in apoptosis and ferroptosis in rectal cancer. Firstly, circABC10 was found to be elevated in rectal cancer tumor tissues and cell lines. After decreased the level of circABC10 in rectal cancer cells using siRNA sequences, and treated with erastin, we found erastin-induced cell death in rectal

cancer cells in a dose-dependent manner and circABC10 depletion significantly increased erastin-induced ferroptosis in rectal cancer cells. At the same time, the obvious accumulation of intracellular lipid ROS and Fe^{2+} was observed in rectal cancer cells. All the results indicated circABC10 depletion induced sensitivity to ferroptosis in rectal cancer cells. Subsequently, circABC10 depletion also was demonstrated to induce cell apoptosis in rectal cancer cells, reflected by the promotion of apoptotic cells and Bax expression, as well as the decreased level of Bcl-2. Besides that, *in vivo* experiments suggested that circABC10 silence inhibited rectal cancer tumor growth *in vivo*.

Both circRNAs and miRNAs have dynamic roles in transcriptional and translational regulation, and interactions between circRNAs and miRNAs are involved in the progression of diverse cancers, including CRC [30, 31]. Thus, we further investigated the potential target miRNAs in binding and regulating circRNAs in rectal cancer. Here, we presented strong evidence showing that circABC10 was a sponge of miR-326 and negatively regulated miR-326 expression. miR-326 is a well-recognized tumor suppressor and associated with the tumorigenesis of CRC [19]. In this

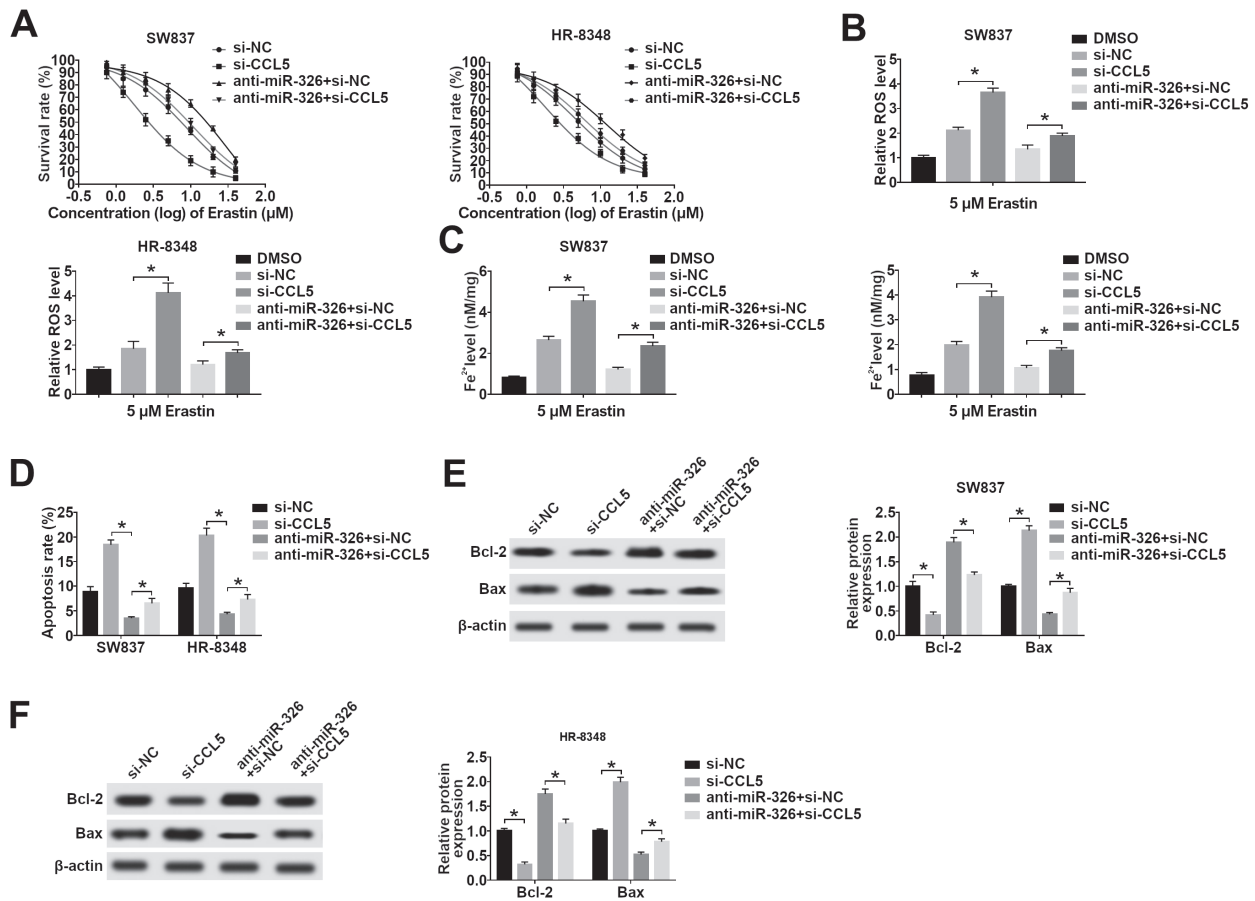


Figure 7. miR-326 inhibition inhibits cell ferroptosis and apoptosis by interacting with CCL5 in rectal cancer. **A)** Cell death was assayed using a CCK-8 kit in SW837 and HR-8348 cells treated with erastin (0–40 μM) for 24 h (One-way ANOVA). **B)** Lipid ROS level was calculated by flow cytometry using C11-BODIPY in SW837 and HR-8348 cells treated with 5 μM erastin for 24 h (One-way ANOVA). **C)** The expression levels of Fe^{2+} accumulation in SW837 and HR-8348 cells treated with 5 μM erastin for 24 h were detected using an iron assay kit (One-way ANOVA). **D)** Cell apoptosis was analyzed using flow cytometry in transfected SW837 and HR-8348 cells (One-way ANOVA). **E, F)** Western blot was used to determine the levels of Bax and Bcl-2 in transfected SW837 and HR-8348 cells (One-way ANOVA). * $p < 0.05$

study, miR-326 was downregulated in rectal cancer, and functional experiments suggested miR-326 performed anti-tumor functions by promoting cell ferroptosis and apoptosis in rectal cancer cell lines. Moreover, rescue assay implied miR-326 inhibition could partially reverse circABC10 depletion-induced promotion on cell ferroptosis and apoptosis in rectal cancer cells.

It is well known that miRNAs play crucial roles in diverse physiological and pathological processes, such as cell proliferation, apoptosis, metastasis, and so on, by directly regulating target gene expression in various tumors [33]. For instance, miR-520 restrained cell proliferation and invasion in CRC by directly interacting with AEG-1 [34]. miR-3174 performed carcinogenic roles in rectal cancer to accelerate cell proliferation and cell cycle by targeting PCBD2 [35]. Here, we next explored the possible target genes of miR-326 using bioinformatics tools, and CCL5 was a target of miR-326 and was negatively regulated by miR-326 in rectal cancer. CCL5 is a

chemotactic cytokine and has been reported to be associated with the development and progression of rectal cancer [36]. In the current study, CCL5 was upregulated in rectal cancer and functioned carcinogenic effects to inhibited cell ferroptosis and apoptosis in rectal cancer cells. Furthermore, we also found CCL5 silencing could partially overturn miR-326 inhibition-induced anti-ferroptosis and anti-apoptosis in rectal cancer cells. Additionally, we also observed that CCL5 was positively regulated by circABC10, and circABC10 served as an endogenous sponge of miR-326 to regulate the CCL5 expression in rectal cancer cells. Thus a circABC10/miR-326/CCL5 regulatory network was identified in rectal cancer cells.

In conclusion, our results demonstrated circABC10 performed carcinogenic functions to suppress ferroptosis and apoptosis in rectal cancer by regulating miR-326/CCL5 axis, suggesting a novel insight for tumor progression and a promising therapeutic target of rectal cancer therapy.

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